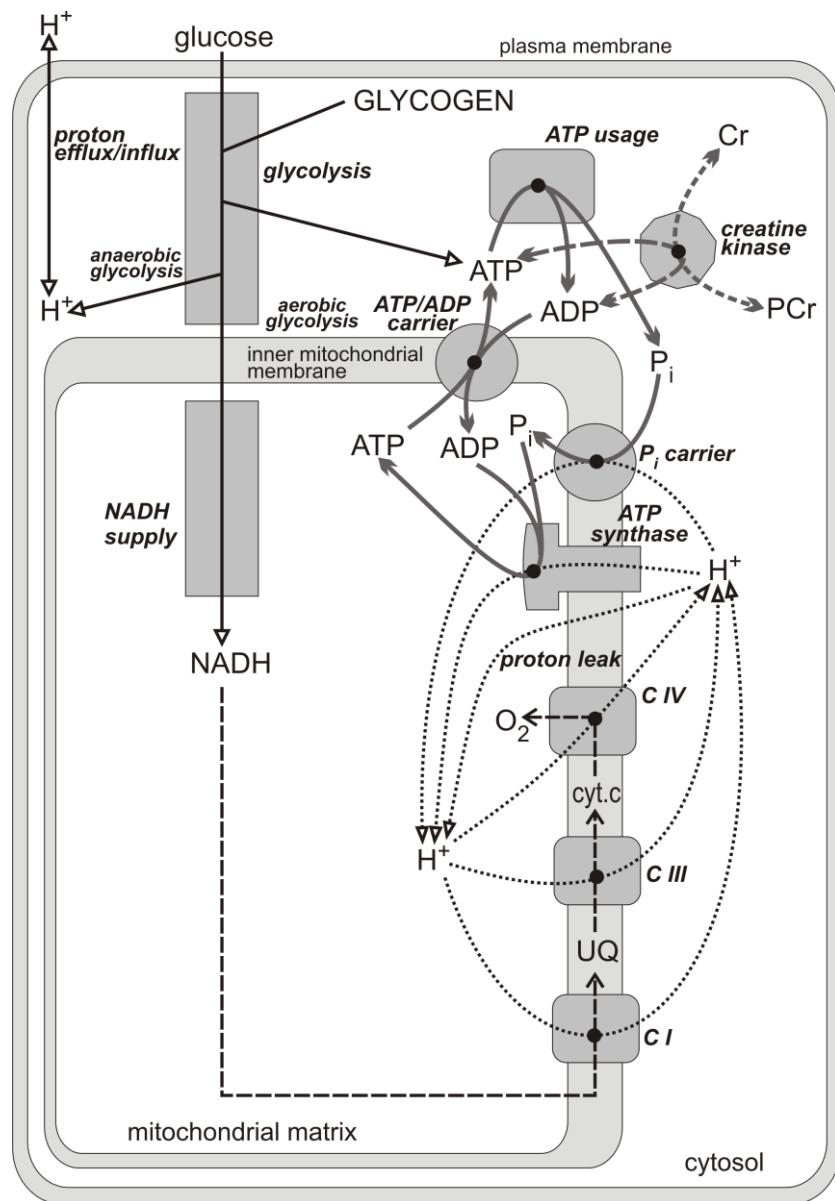


Kinetic description of the dynamic model of the skeletal muscle cell bioenergetic system.



Subscripts: e, external (cytosolic); i, internal (mitochondrial); t, total; f, free; m, magnesium complex; j, monovalent.

All metabolite concentrations in μM . All rates/fluxes in $\mu\text{M min}^{-1}$.

DH, NADH supply; C1, complex I; C3, complex III; C4, complex IV; SN, ATP synthase; EX, ATP/ADP carrier; PI, Pi carrier; UT, ATP usage; LK, proton leak; CK, creatine kinase; AK, adenylate kinase; GL, glycolysis; EF, proton exfflux/influx to/from blood.

Constants

$$k_{DH} = 28074 \mu\text{M min}^{-1}$$

$$K_{mN} = 100$$

$$p_D = 0.8$$

$$k_{C1} = 238.95 \mu\text{M mV}^{-1} \text{ min}^{-1}$$

$$k_{C3} = 136.41 \mu\text{M mV}^{-1} \text{ min}^{-1}$$

$$k_{C4} = 3.600 \mu\text{M}^{-1} \text{ min}^{-1}$$

$K_{mO} = 120 \mu\text{M}$ (mechanistic K_m for O_2 , much higher than apparent K_m)

$$k_{SN} = 34316 \mu\text{M min}^{-1}$$

$n_A = 2.5$ (phenomenological H^+/ATP stoichiometry of ATP syntahse)

$$k_{EX} = 54572 \mu\text{M min}^{-1}$$

$$K_{mADP} = 3.5 \mu\text{M}$$

$$k_{PI} = 69.421 \mu\text{M}^{-1} \text{ min}^{-1}$$

$$k_{UT} = 781.97 \mu\text{M min}^{-1}$$
 (resting state)

$$K_{mA} = 150 \mu\text{M}$$

$$k_{LK1} = 2.500 \mu\text{M min}^{-1}$$

$$k_{LK2} = 0.038 \text{ mV}^{-1}$$

$$k_{fAK} = 862.10 \mu\text{M}^{-1} \text{ min}^{-1}$$

$$k_{bAK} = 22.747 \mu\text{M}^{-1} \text{ min}^{-1}$$

$$k_{fCK} = 1.9258 \mu\text{M}^{-2} \text{ min}^{-1}$$

$$k_{bCK} = 0.00087538 \mu\text{M}^{-1} \text{ min}^{-1}$$

$$k_{EF} = 10000 \mu\text{M min}^{-1}$$

$$\text{pH}_0 = 7.0$$

$$k_{GL} = 17.4 \text{ min}^{-1}$$

$$H^+_{\text{rest}} = 0.1 \mu\text{M}$$

$$K_{DTe} = 24 \mu\text{M}$$
 (magnesium dissociation constant for external ATP)

$$K_{DDe} = 347 \mu\text{M}$$
 (magnesium dissociation constant for external ADP)

$$K_{DTi} = 17 \mu\text{M}$$
 (magnesium dissociation constant for internal ATP)

$$K_{DDi} = 282 \mu\text{M}$$
 (magnesium dissociation constant for internal ADP)

$$R_{Cm} = 15$$
 (cell volume/mitochondria volume ratio)

$$B_N = 5$$
 (buffering capacity coefficient for NAD)

$$T = 298$$

$$R = 0.0083 \text{ kJ mol}^{-1} \text{ K}^{-1}$$

$$F = 0.0965 \text{ kJ mol}^{-1} \text{ mV}^{-1}$$

$$S = 2.303 \cdot R \cdot T$$

$$Z = 2.303 \cdot R \cdot T / F$$

$$u = 0.861 \quad (= \Delta \Psi / \Delta p)$$

$$C_{\text{buffi}} = 0.022 \text{ M H}^+/\text{pH unit} \quad (\text{buffering capacity for H}^+ \text{ in matrix})$$

$$C_{\text{buffe}} = 0.025 \text{ M H}^+/\text{pH unit} \quad (\text{buffering capacity for H}^+ \text{ in cytosol})$$

$$pK_a = 6.8$$

$$\Delta G_{P0} = 31.9 \text{ kJ } \cdot \text{mol}^{-1}$$

$$E_{mN0} = -320 \text{ mV}$$

$$E_{mU0} = 85 \text{ mV}$$

$$E_{mc0} = 250 \text{ mV}$$

$$E_{ma0} = 540 \text{ mV}$$

Constant metabolite concentrations

$$O_2 = 30 \mu\text{M}$$

$$c_t = 270 \mu\text{M} \quad (= c^{2+} + c^{3+}, \text{ total concentration of cytochrome c})$$

$$U_t = 1350 \mu\text{M} \quad (= UQH_2 + UQ, \text{ total concentration of ubiquinone})$$

$$N_t = 2970 \mu\text{M} \quad (= NADH + NAD^+, \text{ total concentration of NAD})$$

$$a_t = 135 \mu\text{M}$$

$$Mg_{fe} = 4000 \mu\text{M} \quad (\text{free external magnesium concentration})$$

$$Mg_{fi} = 380 \mu\text{M} \quad (\text{free internal magnesium concentration})$$

$$A_{iSUM} = 16260 \mu\text{M} \quad (= ATP_{ti} + ADP_{ti}, \text{ total internal adenine nucleotide concentration})$$

$$A_{eSUM} = 6700 \mu\text{M} \quad (= ATP_{te} + ADP_{te} + AMP_e, \text{ total external adenine nucleotide concentration})$$

$$C_{SUM} = 35000 \mu\text{M} \quad (= Cr + PCr, \text{ total creatine concentration})$$

Values of independent variables, respiration rate (v_{C4}) and AMP_e at rest

$$v_{C4} = 287 \mu\text{M min}^{-1}$$

$$NADH = 1669.5 \mu\text{M}$$

$$UQH_2 = 1145.3 \mu\text{M}$$

$$c^{2+} = 53.79 \mu\text{M}$$

$$O_2 = 240.00 \mu\text{M}$$

$$ATP_{ti} = 13580$$

$$Pi_{ti} = 15613 \mu\text{M}$$

$$H_i = 0.03536 \mu\text{M}$$

$$ATP_{te} = 6693.6 \mu\text{M}$$

$$ADP_{te} = 6.599 \mu\text{M}$$

$$(AMP_e = 0.0182) \mu\text{M}$$

$$Pi_{te} = 2823.3 \mu\text{M}$$

$$PCr = 28761 \mu\text{M}$$

$$H_e = 0.1000 \mu\text{M}$$

Calculations

$$C^{3+} = C_t - C^{2+}$$

$$UQ = U_t - UQH_2$$

$$NAD^+ = N_t - NADH$$

$$Cr = C_{SUM} - PCr$$

$$AMP_e = A_{eSUM} - ATP_{te} - ADP_{te}$$

$$ADP_{ti} = A_{iSUM} - ATP_{ti}$$

$$ATP_{fe} = ATP_{te}/(1+Mg_{fe}/k_{DTe})$$

$$ATP_{me} = ATP_{te} - ATP_{fe}$$

$$ADP_{fe} = ADP_{te}/(1+Mg_{fe}/k_{DDe})$$

$$ADP_{me} = ADP_{te} - ADP_{fe}$$

$$ATP_{fi} = ATP_{ti}/(1+Mg_{fi}/k_{DTi})$$

$$ATP_{mi} = ATP_{ti} - ATP_{fi}$$

$$ADP_{fi} = ADP_{ti}/(1+Mg_{fi}/k_{DDi})$$

$$ADP_{mi} = ADP_{ti} - ADP_{fi}$$

$$pH_i = -\log(H_i/10^6) \quad (H_i \text{ expressed in } \mu M)$$

$$pH_e = -\log(H_e/10^6) \quad (H_e \text{ expressed in } \mu M)$$

$$\Delta pH \text{ (mV)} = Z \text{ (pH}_i\text{-pH}_e\text{)}$$

$$\Delta p \text{ (mV)} = 1/(1-u) \Delta pH$$

$$\Delta\Psi \text{ (mV)} = -(\Delta p - \Delta pH)$$

$$\Psi_i \text{ (mV)} = 0.65 \cdot \Delta\Psi$$

$$\Psi_e \text{ (mV)} = -0.35 \cdot \Delta\Psi$$

$$C_{0i} = (10^{-pH_i} - 10^{-pH_i-dpH})/dpH \quad (\text{'natural' buffering capacity for H}^+ \text{ in matrix})$$

$$dpH = 0.001$$

$$r_{buffi} = C_{buffi}/C_{0i} \quad (\text{buffering capacity coefficient for H}^+ \text{ in matrix})$$

$$C_{0e} = (10^{-pH_e} - 10^{-pH_e-dpH})/dpH \quad (\text{'natural' buffering capacity for H}^+ \text{ in cytosol})$$

$$dpH = 0.001$$

$$r_{buffe} = C_{buffe}/C_{0e} \quad (\text{buffering capacity coefficient for H}^+ \text{ in cytosol})$$

$$P_{i,e} = P_{i,e}/(1 + 10^{pH_e-pK_a})$$

$$P_{i,i} = P_{i,i}/(1 + 10^{pH_i-pK_a})$$

$$\Delta G_{SN} = n_A \cdot \Delta p - \Delta G_P \quad (\text{thermodynamic span of ATP synthase})$$

$$\Delta G_P = \Delta G_{P0}/F + Z \cdot \log(10^6 \cdot ATP_{ti}/(ADP_{ti} \cdot P_{i,t})) \quad (\text{concentrations expressed in } \mu M)$$

$$E_{mN} = E_{mN0} + Z/2 \cdot \log(NAD^+/NADH) \quad (\text{NAD redox potential})$$

$$E_{mU} = E_{mU0} + Z/2 \cdot \log(UQ/UQH_2) \quad (\text{ubiquinone redox potential})$$

$$E_{mc} = E_{mc0} + Z \cdot \log(C^{3+}/C^{2+}) \quad (\text{cytochrome c redox potential})$$

$$E_{ma} = E_{mc} + \Delta p \cdot (2 + 2u)/2 \quad (\text{cytochrome a}_3 \text{ redox potential})$$

$$A_{3/2} = 10^{(E_{ma}-E_{ma0})/Z} \quad (a^{3+}/a^{2+} \text{ ratio})$$

$$a^{2+} = a_i/(1 + A_{3/2}) \quad (\text{concentration of reduced cytochrome a}_3)$$

$$\Delta G_{C1} = E_{mU} - E_{mN} - \Delta p \cdot 4/2 \quad (\text{thermodynamic span of complex I})$$

$$\Delta G_{C3} = E_{mc} - E_{mU} - \Delta p \cdot (4-2u)/2 \quad (\text{thermodynamic span of complex III})$$

$$s = 0.7 - (pH - 6.0) \cdot 0.5 \quad (\text{net stoichiometry of proton consumption/production by creatine kinase when coupled with ATP consumption/production, respectively; Lohman reaction})$$

Kinetic equations

Substrate dehydrogenation:

$$v_{DH} = k_{DH} \frac{1}{\left(1 + \frac{K_{mN}}{NAD^+ / NADH}\right)^{p_D}}$$

Complex I:

$$v_{C1} = k_{C1} \cdot \Delta G_{C1}$$

Complex III:

$$v_{C3} = k_{C3} \cdot \Delta G_{C3}$$

Complex IV:

$$v_{C4} = k_{C4} \cdot a^{2+} \cdot c^{2+} \frac{1}{1 + \frac{K_{mO}}{O_2}}$$

ATP synthase:

$$v_{SN} = k_{SN} \frac{\gamma - 1}{\gamma + 1},$$

$$\gamma = 10^{\Delta G_{SN} / Z}$$

ATP/ADP carrier:

$$v_{EX} = k_{EX} \cdot \left(\frac{ADP_{fe}}{ADP_{fe} + ATP_{fe} \cdot 10^{-\Psi_e / Z}} - \frac{ADP_{fi}}{ADP_{fi} + ATP_{fi} \cdot 10^{-\Psi_i / Z}} \right) \cdot \left(\frac{1}{1 + K_{mADP} / ADP_{fe}} \right)$$

Phosphate carrier:

$$v_{PI} = k_{PI} \cdot (Pi_{je} \cdot H_e - Pi_{ji} \cdot H_i)$$

ATP usage:

$$v_{UT} = k_{UT} \frac{1}{1 + \frac{K_{mA}}{ATP_{te}}}$$

Proton leak:

$$v_{LK} = k_{LK1} \cdot (e^{k_{LK2} \cdot \Delta p} - 1)$$

Adenylate kinase:

$$v_{AK} = k_{fAK} \cdot ADP_{fe} \cdot ADP_{me} - k_{bAK} \cdot ATP_{me} \cdot AMP_e$$

Creatine kinase:

$$v_{CK} = k_{fCK} \cdot ADP_{te} \cdot PCr \cdot H_e^+ - k_{bCK} \cdot ATP_{te} \cdot Cr$$

Proton efflux:

$$v_{EF} = k_{EF} \cdot (pH_0 - pH_e)$$

Glycolysis:

$$v_{GL} = k_{GL} \cdot (ADP_{te} + AMP_e) \left(H_{rest}^+ / H^+ \right) \quad (\text{anaerobic glycolysis present})$$

or

$$v_{GL} = 0.2 \cdot v_{DH} \quad (\text{anaerobic glycolysis absent})$$

Set of differential equations

$$\dot{NADH} = (v_{DH} - v_{C1}) \cdot R_{cm} / B_N$$

$$\dot{UQH_2} = (v_{C1} - v_{C3}) \cdot R_{cm}$$

$$\dot{c^{2+}} = (v_{C3} - 2 \cdot v_{C4}) \cdot 2 \cdot R_{cm}$$

$$\dot{O_2} = 0 \quad (\text{constant saturated oxygen concentration} = 240 \mu\text{M}) \text{ or } \dot{O_2} = -v_{C4}$$

$$\dot{H_i^+} =$$

$$-(2 \cdot (2 + 2 \cdot u) \cdot v_{C4} + (4 - 2 \cdot u) \cdot v_{C3} + 4 \cdot v_{C1} - n_A \cdot v_{SN} - u \cdot v_{EX} - (1 - u) \cdot v_{PI} - v_{LK}) \cdot R_{cm} /$$

$$r_{buffi}$$

$$\dot{ATP}_{ti} = (v_{SN} - v_{EX}) \cdot R_{cm}$$

$$\dot{Pi}_{ti} = (v_{PI} - v_{SN}) \cdot R_{cm}$$

$$\dot{ATP}_{te} = (v_{EX} - v_{UT} + v_{AK} + v_{CK} + 1.5 \cdot v_{GL}) \cdot R_{cm} / (R_{cm} - 1)$$

$$\dot{ADP}_{te} = (v_{UT} - v_{EX} - 2 \cdot v_{AK} - v_{CK} - 1.5 \cdot v_{GL}) \cdot R_{cm} / (R_{cm} - 1)$$

$$\dot{Pi}_{te} = (v_{UT} - v_{PI} - 1.5 \cdot v_{GL}) \cdot R_{cm} / (R_{cm} - 1)$$

$$\dot{PCr} = -v_{CK} \cdot R_{cm} / (R_{cm} - 1)$$

$$\dot{H_e^+} =$$

$$\left(2 \cdot (2 + 2 \cdot u) \cdot v_{C4} + (4 - 2 \cdot u) \cdot v_{C3} + 4 \cdot v_{C1} - n_A \cdot v_{SN} - u \cdot v_{EX} - (1 - u) \cdot v_{PI} - v_{LK} - \right) /$$

$$r_{buffe} \cdot R_{cm} / (R_{cm} - 1)$$

Simulations of work transitions (rest-work-rest transitions)

See:

- Korzeniewski B, Zoladz JA.** Slow V'VO₂ off-kinetics in skeletal muscle is associated with fast PCr off-kinetics – and inversely. *J Appl Physiol* 115: 605-612, 2013.
- Korzeniewski B.** Regulation of oxidative phosphorylation during work transitions results from its kinetic properties. *J Appl Physiol* 116: 83-94, 2014.
- Korzeniewski B.** Regulation of oxidative phosphorylation in different muscles and various experimental conditions. *Biochem J* 375: 799-804, 2003.
- Korzeniewski B, Rossiter HB.** Each-step activation of oxidative phosphorylation is necessary to explain muscle metabolic kinetic responses to exercise and recovery in humans. *J Physiol* 593: 5255-5268, 2015.
- Korzeniewski B** (2017) Regulation of oxidative phosphorylation through each-step activation: evidences from computer modeling. *Prog Biophys Mol Biol* 125, 1-23.
- Korzeniewski B** (2018) Regulation of oxidative phosphorylation is different in electrically- and cortically-stimulated skeletal muscle. *PLoS One* 13(4): e0195620. <https://doi.org/10.1371/journal.pone.0195620>.
- Korzeniewski B** (2018) Muscle VO₂-power output nonlinearity in constant-power, step-incremental, and ramp-incremental exercise: magnitude and underlying mechanisms. *Phys Rep* 6(21), e13915. <https://doi.org/10.14814/phy2.13915>.
- Korzeniewski B, Rossiter HB** (2020) Exceeding a "critical" muscle P_i: implications for VO₂ and metabolite slow components, muscle fatigue and power-duration relationship. *Eur J Appl Physiol* 120, 1609-1619. <https://doi.org/10.1007/s00421-0209-04388-4>.
- Korzeniewski B , Rossiter HB** (2021) Factors determining training-induced changes in VO_{2max}, critical power and VO₂ on-kinetics in skeletal muscle. *J Appl Physiol* 130: 498-507. <https://doi.org/10.1152/japplphysiol.00745.2020>
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